

BioControl **46:** 61–70, 2001. © 2001 *Kluwer Academic Publishers. Printed in the Netherlands.*

Status of biological control by egg parasitoids of *Diaprepes abbreviatus* (Coleoptera: Curculionidae) in citrus in Florida and Puerto Rico

D.G. HALL^{1*}, J. PEÑA², R. FRANQUI³, R. NGUYEN⁴, P. STANSLY⁵, C. McCOY⁶, S.L. LAPOINTE⁷, R.C. ADAIR⁸ and B. BULLOCK⁹

¹Research Department, United States Sugar Corporation, Clewiston, Florida; ²Tropical Research & Education Center, University of Florida, Homestead, Florida; ³Agricultural Experiment Station, University of Puerto Rico, Rio Piedras, Puerto Rico; ⁴Division of Plant Industry, Department of Agriculture & Consumer Services, Gainesville, Florida; ⁵Southwest Florida Research & Education Center, University of Florida, Immokalee, Florida; ⁶Citrus Research & Education Center, University of Florida, Lake Alfred, Florida; ⁷USDA-ARS, US Horticultural Research Laboratory, Ft Pierce, Florida; ⁸The Kerr Center for Sustainable Agriculture, Vero Beach Research Station, Vero Beach, Florida; ⁹AREC, University of Florida, Ft Pierce, Florida, USA

*author for correspondence; e-mail: dghall@ussugar.com

Received 8 February 2000; accepted in revised form 13 July 2000

Abstract. Eggs of *Diaprepes abbreviatus* (L.) were routinely monitored in citrus groves at ten locations in Florida during 1997 and 1998 to study egg parasitism. One citrus location was studied in Puerto Rico. No native parasitoids were recovered from 1,337 *D. abbreviatus* egg masses studied in Florida citrus. In contrast, an average of 35.5% (range 12.5 to 68.8%) parasitism of egg masses was reported in Puerto Rico. The parasitoids *Aprostocetus gala*, *Horismenus* spp, and *Quadrastichus haitiensis* were recovered from the eggs of *D. abbreviatus* studied in Puerto Rico. The *Horismenus* parasitoids were suspected hyperparasitoids. Releases of the parasitoid *Ceratogramma etiennei* from Guadeloupe were initiated during 1998 at each of the Florida research sites. By the end of 1998, *C. etiennei* had been recovered from *D. abbreviatus* eggs at two of nine locations in Florida citrus. The parasitoid was recovered from 1 of 34 egg masses at one of these locations during the month of September and from 3 of 34 egg masses at the other location during the month of November. Whether or not *C. etiennei* establishes itself at one or more locations in Florida remains to be seen.

Key words: citrus, classical biological control, egg parasitoids, root weevils, *Ceratogramma etiennei*, *Diaprepes abbreviatus*

Introduction

The root weevil *Diaprepes abbreviatus* (L.) is an important citrus pest in Florida (Hall, 1995; McCoy et al., 1995). Larvae of the weevil can cause substantial damage to roots resulting in decreased production and sometimes

plant death (Woodruff, 1968). Larvae and adults are highly polyphagous (Simpson et al., 1996). Adult female *D. abbreviatus* place their eggs with an adhesive in a single layer in a niche fabricated by juxtaposing leaves (Howden, 1995) or other available surfaces (Wolcott, 1933). Neonates fall to the ground and enter the soil where they feed on progressively larger roots as they grow (Browning et al., 1995). The larvae can infest and destroy the root systems of both seedlings and mature citrus trees (Beavers et al., 1979). Damage to roots is often in the form of girdling, which can cause the death of small citrus trees (Woodruff, 1968) and probably larger trees. In addition to direct damage, larval feeding provides infection sites for plant pathogens, notably *Phytophthora* spp., that can contribute to tree mortality and reductions in productivity (Rogers et al., 1996).

Management strategies currently advocated for an infestation of *Diaprepes abbreviatus* in Florida citrus include chemical sprays for adult and egg control, entomopathogenic nematodes for control of larvae, tactics to control *Phytophthora* if this fungal pathogen is present in an infested grove, and carefully managed tree fertilization and irrigation practices to prevent tree stress (Bullock et al., 1999). A management program employing all of these control tactics is expensive, and general observations indicate that the program may reduce but not prevent economic losses due to the weevil. Additional and alternative management tactics are therefore being researched.

Biological control by egg parasitoids may hold some potential as an additional component of a management program for the weevil in Florida citrus. Although no indigenous egg parasitoids have been reported to attack Diaprepes abbreviatus in Florida, reports and museum specimens from Puerto Rico, Dominican Republic, Dominica and other Caribbean and West Indian countries indicate that five parasitoid species may attack eggs of D. abbreviatus: Ceratogramma etiennei Delvare (Hymenoptera: Trichogrammatidae) in Guadeloupe; Aprostocetus gala (Walker) (= Aprostocetus vaquitarum (Wolcott)) (Hymenoptera: Eulophidae) in Puerto Rico, Guadeloupe, and Dominica; Quadrastichus (previously placed in Tetrastichus) haitiensis Gahan (Hymenoptera: Eulophidae) in Guadeloupe, Puerto Rico, and Dominica; Q. fennahi Schauff (Hymenoptera: Eulophidae) in St Lucia and Barbados; and Brachyufens osborni (Dozier) (Hymenoptera: Trichogrammatidae) in Puerto Rico, Dominican Republic and Andros (Etienne and Delvare, 1991; Schauff, 1987; Pinto and Viggiani, 1991) [questions have been raised regarding whether some of these parasitoid species actually parasitize D. abbreviatus]. The parasitoid Horismenus bennetti Schauff (Hymenoptera: Eulophidae) has been recovered from eggs of D. abbreviatus in the Caribbean and West Indies, but it is thought to be a hyperparasite of *Quadrastichus* spp. (Schauff, 1987). The economic benefits of biological control by parasitoids

of *D. abbreviatus* eggs are not known, however, *D. abbreviatus* is generally considered an important pest in the Caribbean and West Indies in spite of the presence of egg parasitoids.

Two parasitoid species in Florida citrus have been reported to attack eggs of the citrus root weevils Pachnaeus litus and P. opalus: Brachyufens osborni (Baranowski, 1960; Beavers et al., 1980) and Trichogramma sp. (Beavers et al., 1980). Neither of these parasitoids has ever been recovered in Florida from eggs of Diaprepes abbreviatus, which raises questions regarding reports that B. osborni parasitizes D. abbreviatus in Caribbean areas. The parasitoid Quadrastichus haitiensis is recognized as an important biological control agent of D. abbreviatus eggs in Puerto Rico (Wolcott, 1948; Woodruff, 1964). Q. haitiensis was imported from Puerto Rico and released in Florida citrus during 1969. Subsequently, the parasitoid was recovered from one D. abbre*viatus* egg mass from a release location near Apopka (Sutton et al., 1972) and from one P. litus egg mass from a release location near West Palm Beach (Beavers and Selhime, 1975). Further releases of the parasitoid were made during 1970 and 1971, but recovery efforts each year indicated the parasitoid did not establish (Beavers and Selhime, 1975). During 1978 in a citrus grove near Oakhill, Florida, in Volusia county, Q. haitiensis was recovered from egg masses of P. opalus, indicating the parasitoid was established in Florida (Beavers et al., 1980). However, weevil eggs screened in citrus during the 1980s and early 1990s produced no parasitoids (H. Browning, personal communication), suggesting that D. abbreviatus is currently not subjected to any biological control by egg parasitoids in Florida. In view of the potential for further releases of parasitoids from the Caribbean and West Indies, additional attempts at classical biological control of D. abbreviatus eggs in Florida citrus appear justified.

Reported here are the results of a survey conducted to determine whether eggs of *Diaprepes abbreviatus* in Florida citrus are attacked by indigenous parasitoids and, if so, to what extent. Similar research was conducted in Puerto Rico for comparison. Releases of the exotic parasitoid *Ceratogramma etiennei* from Guadeloupe were subsequently initiated in Florida citrus. *C. etiennei* was chosen for releases because co-author Peña had established a colony and had received clearance to release the parasitoid. The outcome of early recovery attempts for this parasitoid is discussed.

Materials and methods

Eggs of *Diaprepes abbreviatus* were monitored for parasitoids during 1997–1998 in Florida citrus at ten locations (eight counties): Moore Haven (Glades Co.); Homestead (Dade Co.); Denaud (Hendry Co.); Vero Beach site #1

| Location | Scion | Rootstock | Soil type | Tree age | Grove size |
|-------------------|-----------------------------|----------------------|----------------------|-----------|---------------|
| Alturas, FL | Hamlin | Swingle | Astatula fine sand | 9–10 yrs | 4 ha |
| Apopka, FL | Hamlin | Carrizo | Sandy loam | 3 yrs | 1 ha |
| Denaud, FL | Mandarins, sweet oranges | Mixed | Pinada Fine Sand | Mixed | 25 ha |
| Ft Pierce, FL #1 | Red grapefruit | Cleopatra | Winder loamy sand | 11 yrs | 32 ha |
| Ft Pierce, FL #2 | Hamlin | Carrizo | Immokalee sand | 6 yrs | 12 ha |
| Homestead, FL | Lime | Wide collection | Rockdale | 1–25 yrs | 1 ha |
| Plant City, FL | Hamlin | Cleopatra Swingle | Orlando fine | 5–12 yrs | 5 ha |
| Moore Haven, FL | Fallglo, Valencia | Swingle | Immokolee sand | 6–7 yrs | 1 ha |
| Vero Beach, FL #1 | White, red grapefruit | Cleopatra Swingle | Winder fine sand | 11–29 yrs | 3 ha |
| Vero Beach, FL #2 | Navels, red grapefruit | Sour orange | Winder fine sand | 8–25 yrs | 4 ha |
| Isabela, PR | Valencia | Cleopatra | Cotto | 3–4 yrs | 2 ha |

Table 1. Citrus sites sampled during the study

(Indian River Co.); Vero Beach site #2 (Indian River Co.); Ft Pierce site #1 (St Lucie Co.); Ft Pierce site #2 (St Lucie Co.); Apopka (Orange Co.); Alturas (Polk Co.); and Plant City (Hillsborough Co.). One location was studied near Isabela in Puerto Rico during 1998–1999. Table 1 presents basic information on the study sites. No insecticides were applied during the study period at any of the locations researched.

The parasite recovery methodology was to, at least once a month, cage ovipositing female weevils onto bouquets of citrus leaves for 24–48 h. The caged bouquets of leaves usually consisted of both mature and expanded flush leaves. One bouquet of leaves was caged per tree within the basal third of the canopy, with up to five trees receiving a cage across an area of 50 to 100 trees (about 0.25 ha). One to five wild adult weevils of each sex were released into each cage. After the 24–48 h period, cages and adults were removed and

leaves with eggs were tagged. Four days later, these leaves with eggs were excised and taken to a laboratory. Individual egg masses (still intact on leaf tissue) were placed into a 30 ml glass vial with a screw cap, and then the vials were placed into a humidified plastic bag on a lab bench under ambient conditions (among the laboratories holding eggs, mean 24.4 °C, SEM 0.57, maximum 26 °C and minimum 22 °C). One week later the caps were removed from the vials, the opening of each vial was plugged with cotton wrapped in a piece of tissue paper, and then the vials were placed back onto a bench under ambient conditions. The vials were examined for neonate larvae and emerged parasites every 4 to 7 days over a 28-day period, during which time the cotton plugs were kept moist. Number of eggs per mass, percent egg hatch, and percent eggs parasitized were determined. For reporting purposes, data from 2 sites sampled near Vero Beach were combined and data from 2 sites near Ft Pierce were combined.

Releases of Ceratogramma etiennei were initiated in Florida during May 1998. The parasitoid was imported from Guadeloupe by co-author J. Peña and reared at Homestead (Peña et al., 1998) and Gainesville. Peña planned a separate publication to review specific rearing procedures for C. etiennei. Briefly, the general methods employed by Beavers and Selhime (1975) to propagate Quadrastichus haitiensis were used to propagate C. etiennei. Adult Diaprepes abbreviatus, field-collected from limes and silver buttonwood (Conocarpus erectus L. variety sericeus Fors. ex DC (Combretaceae)), were held in a laboratory for oviposition on wax paper. A number of strips of wax paper with fresh eggs were then exposed to adults of C. etiennei (generally at a ratio of 2 to 8 masses per female parasitoid) in a glove box. All rearing was conducted at 24-26 °C, 12:12 L:D photoperiod, 75-81% RH. Progeny were field-released either by hanging wax paper strips with nearly-mature parasitoid pupae in trees, by placing wax paper strips with nearly-mature parasitoid pupae in paper emergence bags hung in citrus trees, or by releasing newly-emerged adults into trees. In cases where adults were released, they were aspirated and released (no honey or other adult food was supplied to the newly-emerged adults). Releases of C. etiennei were made within the same 50- to 100-tree areas in which we surveyed for parasitoids of D. abbreviatus eggs, and the releases sometimes coincided with periods of time during which we were sampling for egg parasitoids (i.e., weevil eggs on tagged leaves were available to parasitoids).

Parasitoid specimens observed during the study were identified by Dr John Pinto (Department of Entomology, University of Riverside, Riverside, CA) and Dr Michael Schauff (USDA-ARS, c/o National Museum Natural History, Washington, DC).

Results and discussion

Totals of 1,337 and 91 egg masses oviposited on citrus leaves by Diaprepes abbreviatus in Florida and Puerto Rico, respectively, were screened for parasitoids during the study (Table 2). A mean \pm SEM of 53.8 \pm 2.8 eggs per mass was observed in Florida and 49.8 ± 3.0 eggs per mass was observed in Puerto Rico (Table 2). Using our holding method in the lab, percentage egg eclosion per mass averaged $62.7 \pm 3.7\%$ among masses studied in Florida and $68.9 \pm 7.9\%$ among masses studied in Puerto Rico (Table 2). Percentages of eclosion per mass lower than 10% were occasionally observed. In cases of low percent hatch, the eggs usually looked normal at first, then turned milky white and later became brown and shriveled. In these cases it was possible that unfertile eggs had been laid, but this was not confirmed. No native parasitoids of D. abbreviatus eggs were recovered in Florida citrus; in contrast, a mean of $35.5 \pm 11.9\%$ parasitism of egg masses (range 12.5 to 68.8%) was reported in Puerto Rico (Table 2). With respect to parasitized masses observed in Puerto Rico, data were not collected on the percentage of parasitized eggs per mass. Aprostocetus gala, Horismenus spp., and Quadrastichus haitiensis were recovered from eggs of D. abbreviatus in Puerto Rico (identifications from co-author R. Franqui), but information on the relative abundance of these parasitoids was not recorded. However, samples of eggs thought to be D. abbreviatus from citrus (egg masses of this weevil morphologically resemble eggs of other citrus root weevil species) were obtained from Puerto Rico during the summer 1998, and parasitoid species recovered under quarantine in Gainesville, Florida, included predominantly Q. haitiensis but also A. gala, Brachyufens osborni, and Horismenus bennetti. As mentioned earlier, H. bennetti was a suspected hyperparasitoid. Because B. osborni in Florida citrus is known to attack Pachneus spp. but not D. abbreviatus, the presence of this parasitoid in the samples suggested either that Puerto Rican populations of B. osborni do parasitize D. abbreviatus in citrus or that some of the weevil egg masses studied were incorrectly identified as D. abbreviatus from citrus. Whether or not B. osborni parasitizes D. abbreviatus in citrus in the Caribbean area needs confirmation.

Over 62,000 adult *Ceratogramma etiennei* were released into Florida citrus during May–November 1998 (Table 3). The number of parasitoids released at a time varied and was dependent upon production. An average total of approximately 1,430 parasites (equivalent to about 5,720 parasites per ha) was released each month at each study location. Individual releases ranged from the equivalent of around 560 to around 1,200 parasites per ha. Usually 1 or 2 individual releases of the parasitoid were made monthly at each study site. A total of 1,181 *Diaprepes abbreviatus* egg masses were screened for parasitoids after releases were initiated, among which *C. etiennei*

| Year | Loca | Variable ^b | Jan | Feb | Mar | Apr | May | Jun | Jul | Aug | Sep | Oct | Nov |
|------|------|-----------------------|----------------|-------|-----|-----|-----|-----------------|----------------|-----|---------|-----|----------------|
| 1997 | Ap | Egg masses | _ | _ | _ | _ | _ | 18 | 9 | 4 | 3 | _ | _ |
| | | Masses parasitized | _ | _ | _ | _ | _ | 0 | 0 | 0 | 0 | _ | _ |
| | Dn | Egg masses | _ | _ | _ | _ | _ | 1 | 30 | 51 | 49 | 53 | 6 |
| | | Masses parasitized | _ | _ | _ | _ | _ | 0 | 0 | 0 | 0 | 0 | 0 |
| | Hm | Egg masses | _ | _ | _ | _ | _ | _ | _ | _ | _ | 1 | _ |
| | | Masses parasitized | | _ | _ | _ | _ | _ | _ | _ | _ | 0 | _ |
| | FP | Egg masses | _ | _ | _ | _ | _ | _ | _ | 20 | 27 | 19 | _ |
| | | Masses parasitized | _ | _ | _ | _ | _ | _ | _ | 0 | 0 | 0 | _ |
| | MH | Egg masses | _ | _ | _ | 11 | 7 | 31 | 7 | 16 | 14 | 5 | 5 |
| | | Masses parasitized | _ | _ | _ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 1998 | Al | Egg masses | _ | _ | 1 | 5 | 5 | _ | 18 | 1 | 52 | _ | _ |
| | | Masses parasitized | | _ | 0 | 0 | 0 | _ | 0 | 0 | 0 | _ | _ |
| | Ap | Egg masses | _ | _ | _ | 21 | 17 | 16 | 8 | 9 | 14 | 3 | _ |
| | | Masses parasitized | _ | _ | _ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | _ |
| | Dn | Egg masses | _ | _ | _ | 11 | 13 | _ | _ | _ | _ | _ | 34 |
| | | Masses parasitized | | _ | _ | 0 | 0 | _ | _ | _ | _ | _ | 3 ^c |
| | FP | Egg masses | _ | _ | _ | _ | 53 | 66 | 51 | 28 | 48 | 30 | 7 |
| | | Masses parasitized | | _ | _ | _ | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | Hm | Egg masses | _ | _ | 2 | 2 | 16 | 5 | _ | 6 | _ | 8 | _ |
| | | Masses parasitized | _ | _ | 0 | 0 | 0 | 0 | _ | 0 | _ | 0 | _ |
| | MH | Egg masses | _ | _ | _ | 8 | 11 | 6 | 5 | 16 | 34 | 13 | 7 |
| | | Masses parasitized | _ | _ | _ | 0 | 0 | 0 | 0 | 0 | 1^{c} | 0 | 0 |
| | PC | Egg masses | _ | _ | 1 | 6 | 3 | 1 | 14 | 4 | 31 | _ | 11 |
| | | Masses parasitized | | _ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | _ | 0 |
| | VB | Egg masses | _ | _ | _ | 29 | 15 | 23 | 31 | 47 | 35 | 21 | 28 |
| | | Masses parasitized | | _ | _ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | PR | Egg masses | _ | _ | _ | _ | _ | 32 | 12 | _ | _ | _ | 11 |
| | | Masses parasitized | _ | _ | _ | _ | _ | 22 ^d | 4 ^d | _ | _ | _ | 3 ^d |
| 1999 | PR | Egg masses | 20 | 16 | _ | _ | _ | _ | _ | _ | _ | _ | _ |
| | | Masses parasitized | 5 ^d | 2^d | — | — | — | _ | — | _ | — | _ | _ |

Table 2. Results of the survey in citrus for parasitoids of *D. abbreviatus* eggs in Florida and Puerto Rico (a dash indicates no sampling was conducted)

^aLocations: Al=Alturas, Ap=Apopka, Dn=Denaud, FP=Ft Pierce, Hm=Homestead,

MH=Moore Haven, PC=Plant City, VB=Vero Beach, PR=Puerto Rico.

^bVariable: Egg masses is number of masses observed, Masses parasitized is number of masses parasitized.

^cParasitoids identified as *Ceratogramma etiennei*.

^dParasitoids identified as *Quadrastichus haitiensis*, *Aprostocetus gala* and *Horismenus* spp.

was recovered from one egg mass at Moore Haven during September and from three egg masses at Denaud during November (Table 2). With respect to the parasites recovered at Moore Haven, the egg mass parasitized had been present on a tree during 16–21 September and the closest *C. etiennei* release date was 26 August. Since adult *C. etiennei* live for 5 to 7 days with perhaps a maximum of 10 days (Amalin et al., unpublished), adult progeny from a previous release (perhaps the one on 26 August) probably parasitized this weevil egg mass at Moore Haven. This particular mass consisted of 42 eggs, all had turned brown a week after removing them from the tree, and 24

| Location | May | Jun | Jul | Aug | Sep | Oct | Nov |
|-------------|-------|-------|-------|-------|-------|-------|-------|
| Alturas | 1,000 | 500 | 1,000 | 500 | 1,000 | 500 | 1,000 |
| Apopka | | _ | 300 | 700 | | 500 | |
| Denaud | 400 | | 2,000 | 1,800 | 7,450 | 2,100 | |
| Homestead | 440 | 450 | 5,900 | 5,900 | 3,900 | 900 | 2,650 |
| Ft Pierce | 140 | 1,330 | 2,300 | 450 | 800 | 500 | |
| Moore Haven | _ | 130 | 540 | 2,200 | | 4,800 | 1,000 |
| Plant City | 1,000 | 500 | 1,000 | 500 | 1,000 | 500 | 1,000 |
| Vero Beach | _ | 250 | — | 260 | 1,070 | 725 | _ |
| | | | | | | | |

Table 3. Releases of *Ceratogramma etiennei*, approximate numbers of adults deployed (1998)^a

^aMean of 73.3% (s.e. 1.75) of the adults were female based on 46 determinations during 1998.

produced parasites a week later (the remaining eggs shriveled). At Denaud, the parasitized egg masses had been present on trees on a *C. etiennei* release date (no data were collected from these masses with respect to numbers of eggs parasitized).

Based on the low incidence of C. etiennei recoveries during 1998, whether or not the parasitoid was establishing at any of the release sites in Florida could not be ascertained. Attempts to establish C. etiennei in the Dominican Republic failed (McCoy, unpublished information). Why Quadrastichus haitiensis and C. etiennei are well established in some Caribbean/West Indian areas and yet may fail to establish in other areas is not known. Beavers and Selhime (1975) speculated that differences in environmental conditions between Puerto Rico and Florida may have contributed to the failure of Q. haitiensis to establish on D. abbreviatus in Florida. Similarly, differences in climate may have contributed to the failure of C. etiennei from Guadeloupe to establish in the Dominican Republic and its possible failure in Florida. Other factors may influence the establishment of parasitoids for biological control of D. abbreviatus eggs in Florida including the availability of food sources (e.g., nectar or pollen) for adult parasites, the particular plant species on which weevils oviposit, and the seasonal availability of host eggs. In any case, Q. haitiensis from Puerto Rico and C. etiennei from Guadeloupe have now each been reared successfully on Florida populations of D. abbreviatus in the laboratory, indicating host compatibility.

Permanent establishment of *C. etiennei* or other egg parasitoids may be achievable in at least some areas in Florida, but whether or not egg parasitoids would reduce the economic importance of *D. abbreviatus* in citrus is not known. Unfortunately, information is lacking on the impact these parasitoids

68

may have against the economic importance of the weevil in the Caribbean and West Indies. The likelihood that egg parasitoids would be of major value in controlling *D. abbreviatus* in Florida is reduced given the fact that the weevil is considered to be an important pest in areas where egg parasitoids are already present (although it is possible in some of these areas that hyperparasitoids confound the benefits of the egg parasitoids to some extent).

Acknowledgments

We thank Dr Michael Schauff and Dr John Pinto for identifying specimens during this project. Dr Schauff also helped clarify scientific nomenclature of the parasitoids. This project was partially funded by the Florida Citrus Production Research Advisory Council, FCPRAC No. 981-58E.

References

- Baranowski, R.M., 1960. Notes on a parasite of the citrus root weevil, *Pachnaeus litus* Germ. *Fla. Entomol.* 43: 197.
- Beavers, J.B. and A.G. Selhime, 1975. Further attempts to establish the weevil egg parasite, *Tetrastichus haitiensis* in Florida. *Fla. Entomol.* 58(1): 29–31.
- Beavers, J.B., R.E. Woodruff, S.A. Lovestrand and W.J. Schroeder, 1979. Bibliography of the sugarcane rootstalk borer weevil, *Diaprepes abbreviatus*. *Bull. Entomol. Soc. Am.* 25: 25–29.
- Beavers, J.B., S.A. Lovestrand and A.G. Selhime, 1980. Establishment of the exotic parasite *Tetrastichus haitiensis* (Hymenoptera: Eulophidae) and recovery of a new Trichogramma (Hymenoptera: Trichogrammatidae) from root weevil egg masses in Florida. *Entomophaga* 25: 91–94.
- Browning, H.W., R.J. McGovern, L.K. Jackson, D.V. Calvert and W.F. Wardowski, 1995. Florida Citrus Diagnostic Guide. Florida Science Source, Inc., Lake Alfred, Florida, USA. 244 pp.
- Bullock, R.C., S.H. Futch, J.L. Knapp and C. W. McCoy, 1999. Citrus root weevils. Fact Sheet ENY-611. In: 1999 Florida Citrus Pest Management Guide, Florida Coop. Ext. Service, University of Florida, Gainesville. SP-43.
- Etienne, J. and G. Delvare, 1991. Les parasites de *Diaprepes abbreviatus* (Coleoptera: Curculionidae) aux Antilles françaises. *Bull. Soc. Entomol. France* 96: 295–299.
- Hall, D.G., 1995. A revision to the bibliography of the sugarcane rootstalk borer weevil, *Diaprepes abbreviatus* (Coleoptera: Curculionidae). *Fla. Entomol.* 78: 364–377.
- Howden, A.T., 1995. Structures related to oviposition in Curculionoidea. Mem. Entomol. Soc. Wash. 14: 53–100.
- McCoy, C.W., E.D. Quintela, S.E. Simpson and J. Fojtik, 1995. Effect of surface-applied and soil-incorporated insecticides for the control of neonate larvae of *Diaprepes abbreviatus* in container-grown citrus. *Proc. Fla. State Hort. Soc.* 108: 130–136.
- Peña, J.E., J. Etienne, R. Duncan and J. Pinto, 1998. Introduction of *Ceratogramma etiennei* (Hymenoptera: Trichogrammatidae) for biological control of *Diaprepes abbreviatus* in

Florida. In: S. Hassan (ed), *Egg Parasitoids*. 5th International Symposium IOBC, Cali, Colombia, March 1998, Parey Buchverlag, Berlin.

- Pinto, D.J. and G. Viggiani, 1991. A taxonomic study of the genus Ceratogramma (Hymenoptera: Trichogrammatidae). Proc. Entomol. Soc. Washington 93: 719–732.
- Rogers, S., J.H. Graham and C.W. McCoy, 1996. Insect-plant pathogen interactions: Preliminary studies of *Diaprepes* root weevil injuries and *Phytophthora* infections. *Proc. Fla. State Hort. Soc.* 109: 57–62.
- Schauff, M.E., 1987. Taxonomy and identification of the egg parasites (Hymenoptera: Platygasteridae, Trichogrammatidae, Mymaridae and Eulophidae) of citrus root weevils (Coleoptera: Curculionidae). *Proc. Entomol. Soc. Washington* 89: 31–42.
- Simpson, S.E., H.N. Nigg, N.C. Coile and R.A. Adair, 1996. *Diaprepes abbreviatus* (Coleoptera: Curculionidae): host plant associations. *Environ. Entomol.* 25: 333–349.
- Sutton, R.A., A.G. Selhime and W. McCloud, 1972. Colonization and release of *Tetrastichus haitiensis* as a biological control agent for citrus root weevils. *J. Econ. Entomol.* 65: 184–185.
- Wolcott, G.N., 1933. Otiorhynchids oviposit between paper. J. Econ. Entomol. 26: 1172-1173.
- Wolcott, G.N., 1948. The insects of Puerto Rico: Coleoptera. J. Agric. Univ. Puerto Rico 32: 225–416.
- Woodruff, R.E., 1964. A Puerto Rican weevil new to the United States (Coleoptera: Curculionidae). *Fla. Dept. Agric., Div. Plant Ind., Entomol. Circ.* 30: 2 pp.
- Woodruff, R.E., 1968. The present status of a West Indian weevil (*Diaprepes abbreviatus* (L.)) in Florida (Coleoptera: Curculionidae). *Fla. Dept. Agric., Div. Plant Ind., Entomol. Circ.* 77: 4 pp.